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# **Cultural practices can reduce damages by *Dryocosmus kuriphilus* in chestnut stands**

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# ***Dryocosmus kuriphilus* arrived in Europe in 2002, in Northern Italy**

**Quarantine not effective**

**Rapid spread in all Italian *Castanea sativa* stands (in 10 years) due to:**

- adult dispersal by flight**
- movement of plants and scions containing eggs**

**The wasp has more recently spread in adjacent countries and has now arrived in Spain**



***Castanea sativa* stands in Italy, about 10% of national forests (INFC, 2007)**



**Infested stands in 2010 (CABI, 2011)**



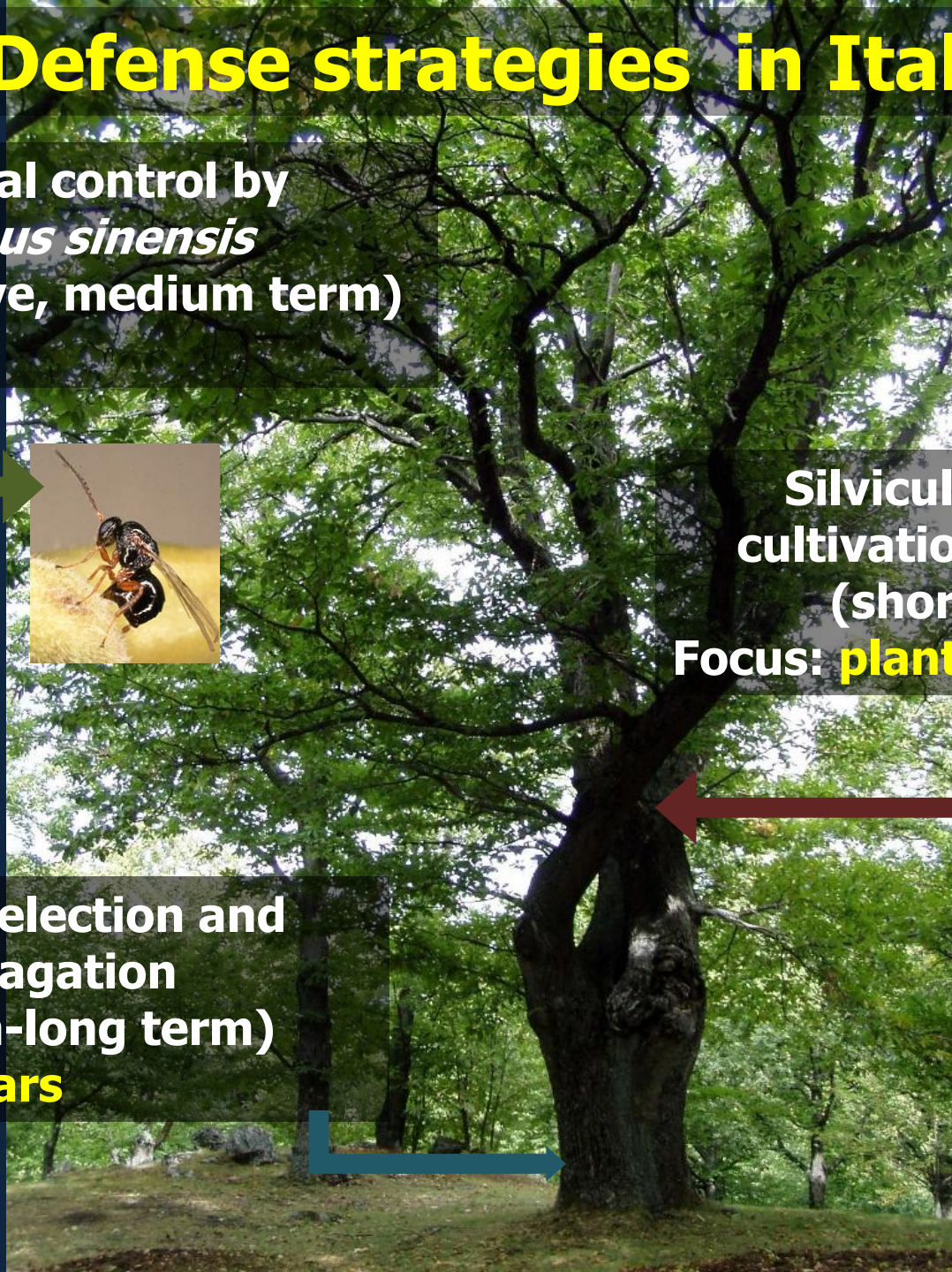
# Defense strategies in Italy

Biological control by  
*Torymus sinensis*  
(best effective, medium term)  
Focus: **insect**



Silvicultural and  
cultivation practices  
(short term)  
Focus: **plants in field**

Genetic selection and  
propagation  
(medium-long term)  
Focus: **cultivars**







**Damage: galls  
on leaves,  
shoots, buds  
and flowers**



**main effect**

**Reduction  
of photosynthetic  
surface**



**Loss of fruit production in  
terms of quantity and quality**

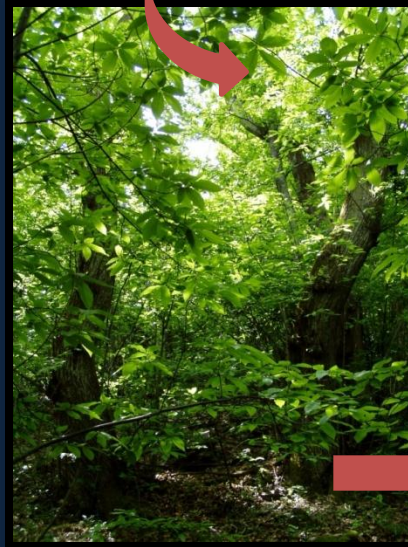
**Reduction of plant growth**



Estimated **loss** in fruit  
production ???  
**50-70%** (CABI, 2011)

**Abandonment of  
cultivation** of  
chestnut orchards

**High costs of  
restoring**  
(as happened after  
chestnut blight)



???

€

During the time gap to the effectiveness of biological control and genetic improvement, it is necessary to reduce the damage to avoid chestnut stands degradation and abandonment



## **Goal**

**to maintain a good vegetative state of plants and an acceptable level of fruit production**

## **Preliminary steps**

**To study the plant-insect interactions**

**Classification of damage  
in types (in terms of consequences  
on plant development)**

**Analysis of the damage types  
distribution on the plant**

**Relationships between  
damage susceptibility and plant vigor**

**(Maltoni *et al.*, 2012)**

# Damage classification

- **POSITION** of the gall (vegetative organ: **Shoot, Leaf, Dormant Bud, Flower**)
- **EFFECT** of the gall on development (**degree of deformity**)



**Abnormal organ development is directly related to a reduction of photosynthetic area causing different consequences according to the attacked organ**



# Galls on leaves and stipules



## Consequences on plant growth

Current season

No

Following seasons

No

Very  
Very  
slight

No

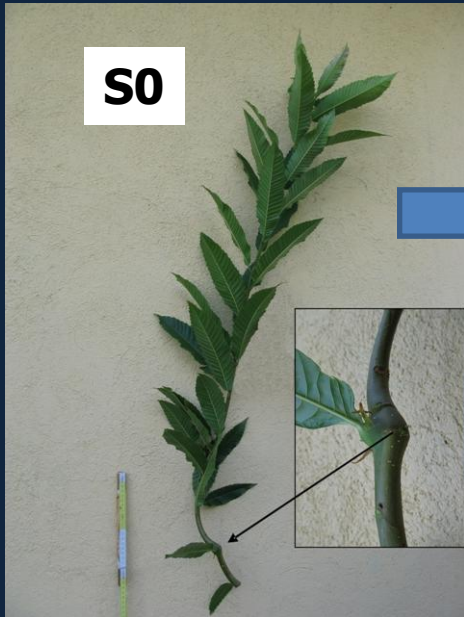
Very  
slight

No



# Galls on shoots

**S0**



The galls are located along the axis or on adjacent leaves and the axis development is nearly normal



**S1**



The galls are located along the axis; deviation of the axis direction and reduction of axis growth and diameter are evident

**S2**



The galls cause a complete deformation in shoot development

# Galls on shoots

S0



**Current  
season**

**Normal shoot growth  
and development**

**Usually this damage  
causes a general  
reduction of the  
active photosynthetic area  
during the current  
growing season**

**Heavy damage**

**It compromises the  
shoot development and  
so the photosynthetic  
activity**

**Following  
seasons**

**No**

**Not predictable (in  
many cases it doesn't  
cause the death of the  
entire shoot)**

**Possible reduction of  
new shoots**

**Most severe damage**

**It always causes the  
shoot death**

**No new shoots**

S1



S2





# Galls on dormant buds and flowers



**DB2**



The gall causes a complete deformation of the bud



**F2**

The gall causes a deformation of the female or male flowers



# Galls on dormant buds and flowers

## Consequences on plant growth

### Current season

### Following seasons

No damage  
(no shoots development  
even in case of normal  
conditions)

No damage in normal  
conditions  
  
Heavy damage if the  
development  
of shoots is required in  
future

Heavy damage  
It involves potential  
fruit production

None

DB2

F2



# Relevance of damage position on shoots



## The development of chestnut shoots



# Rough assessment of the reduction of photosynthetic surface in terms of leaves number

Apical part of the shoot



new shoots

Lower part of the shoot

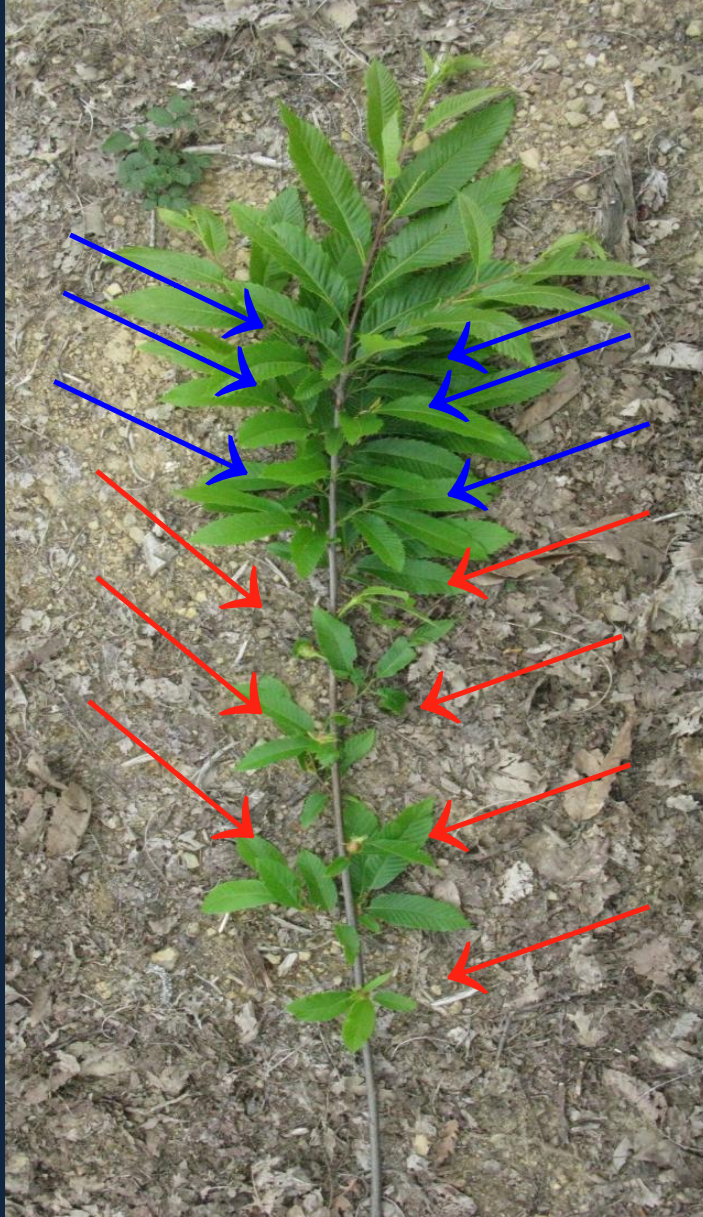


brachiblasts

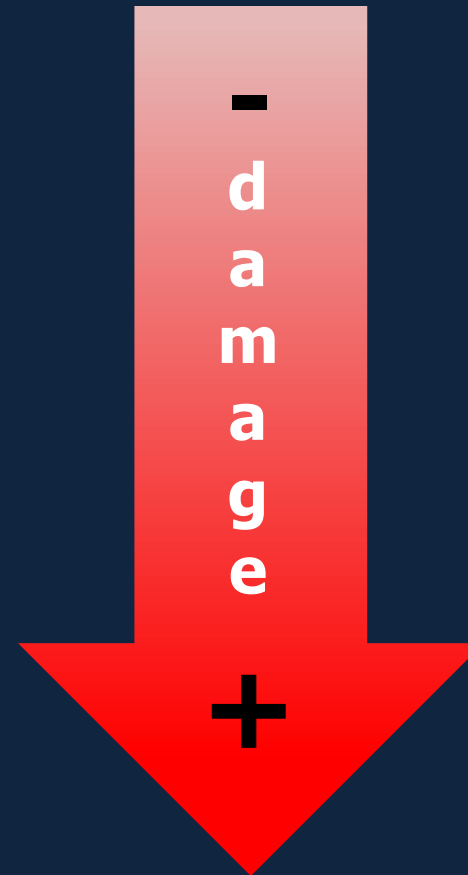
	$Y_0$	$Y_1$	$Y_2$	.... $Y_{10}$	$Y_0$	$Y_1$	$Y_2$	.... $Y_{10}$
St0	0	0	0	0	0	0	0	0
L0	0	0	0	0	0	0	0	0
L1	1	0	0	0	1	0	0	0
L2	1	0	0	0	1	0	0	0
S0	0	0	0	0	0	0	0	0
S1	0-5	0-30	0-90	0-3000	0-3	0-3	0-3	0-3
S2	10	~35	>100	~4500	3	3	3	3
DB2					0	0-?	0-?	
F2	5	0	0	0	5	0	0	0



# Analysis of the damage types distribution on the plant



**Damage distribution on shoots is not random**



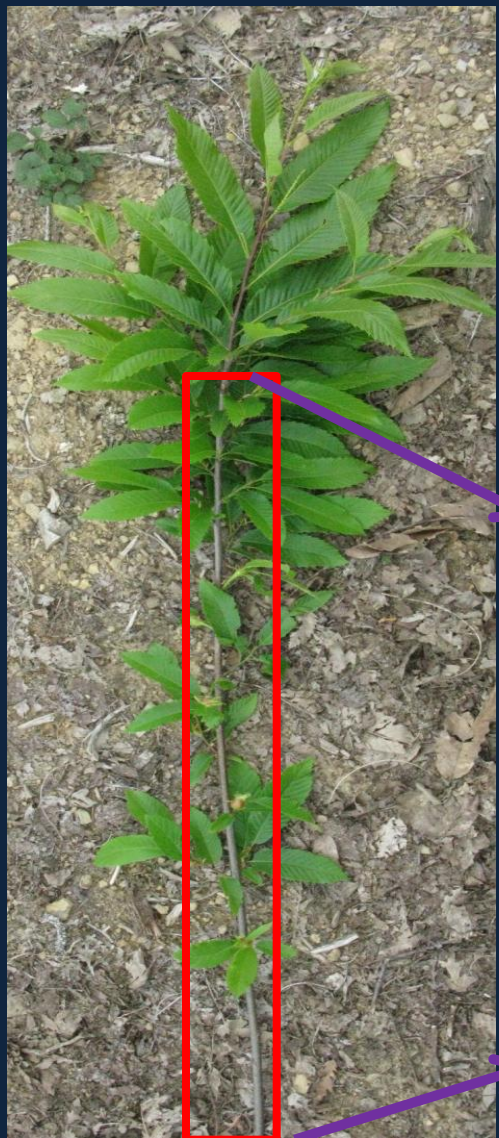
(Maltoni et al., 2012)

# **Relationships between damage susceptibility and plant vigor**

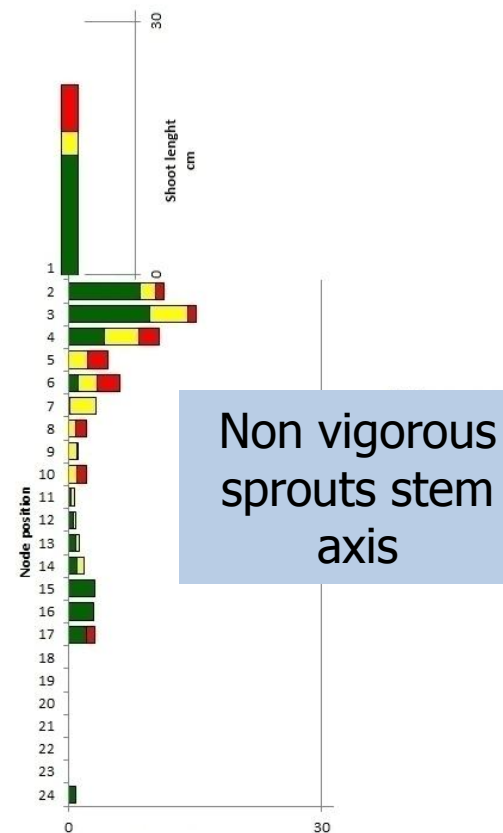
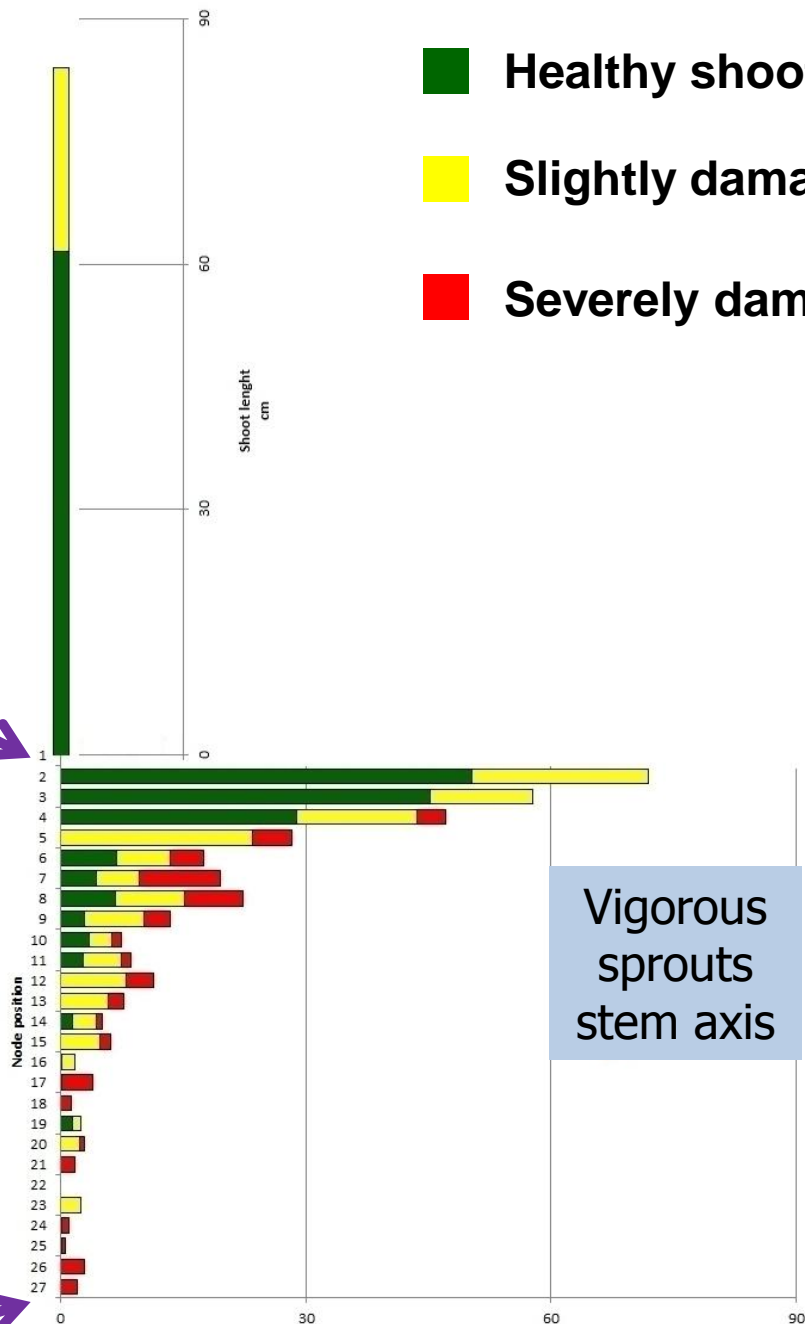
**Why is it important to investigate it?**

**If there is a relationship between the damage and plant vigor, it is possible to experiment with effective cultural practices focused on reducing the damages on the plant**



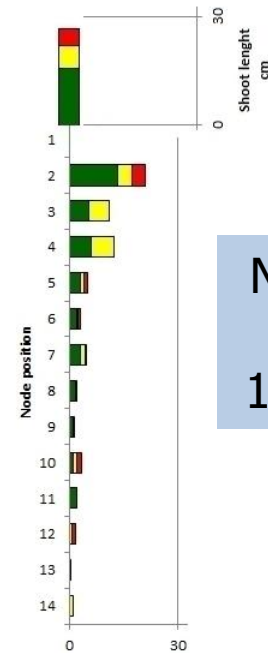
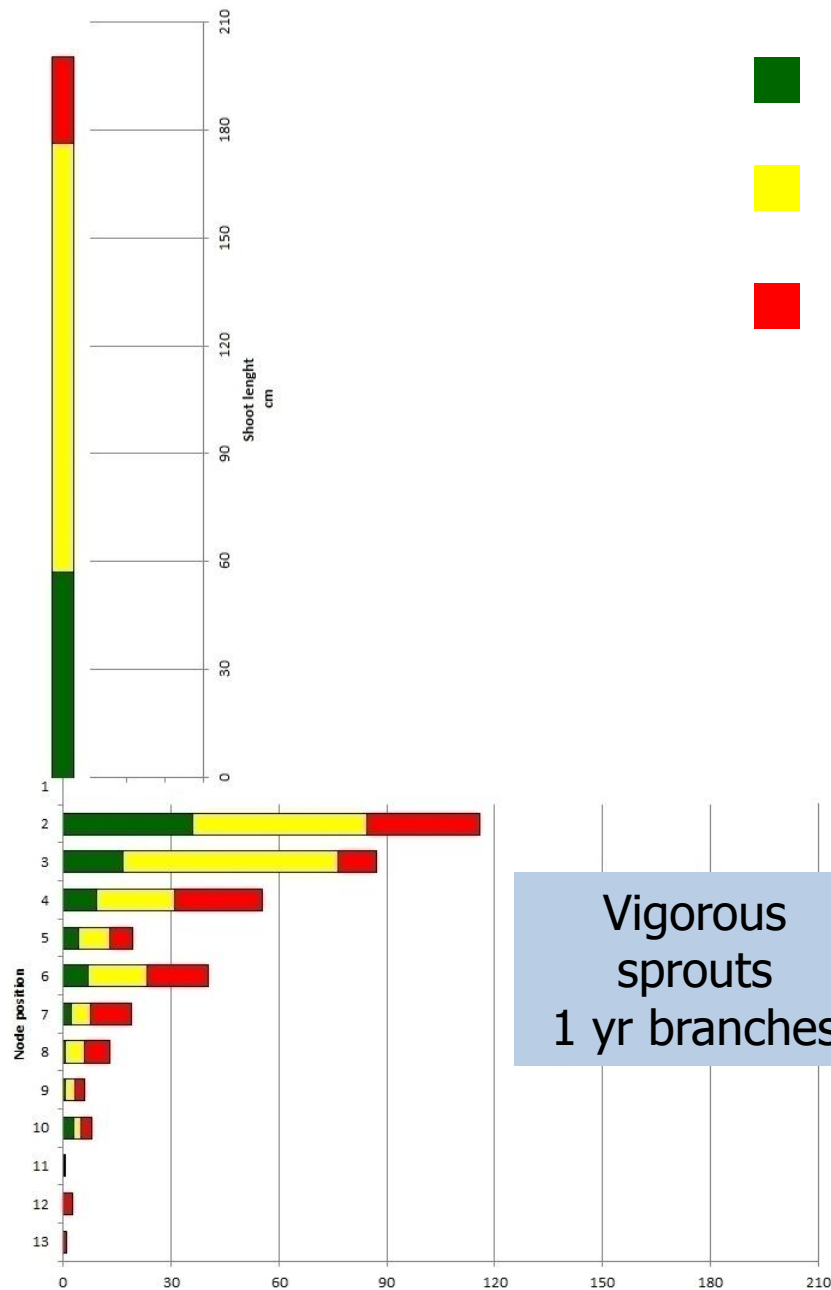


- Healthy shoots
- Slightly damaged shoots
- Severely damaged shoots



Shoots lenght sum (cm)

- Healthy shoots
- Slightly damaged shoots
- Severely damaged shoots



Shoots length sum (cm)



A photograph of a plant shoot against a clear blue sky. The shoot is dark brown and has several green, serrated leaves. A green circle highlights the upper, more developed part of the shoot, while a red circle highlights a lower section where the leaves are smaller and the shoot appears damaged.

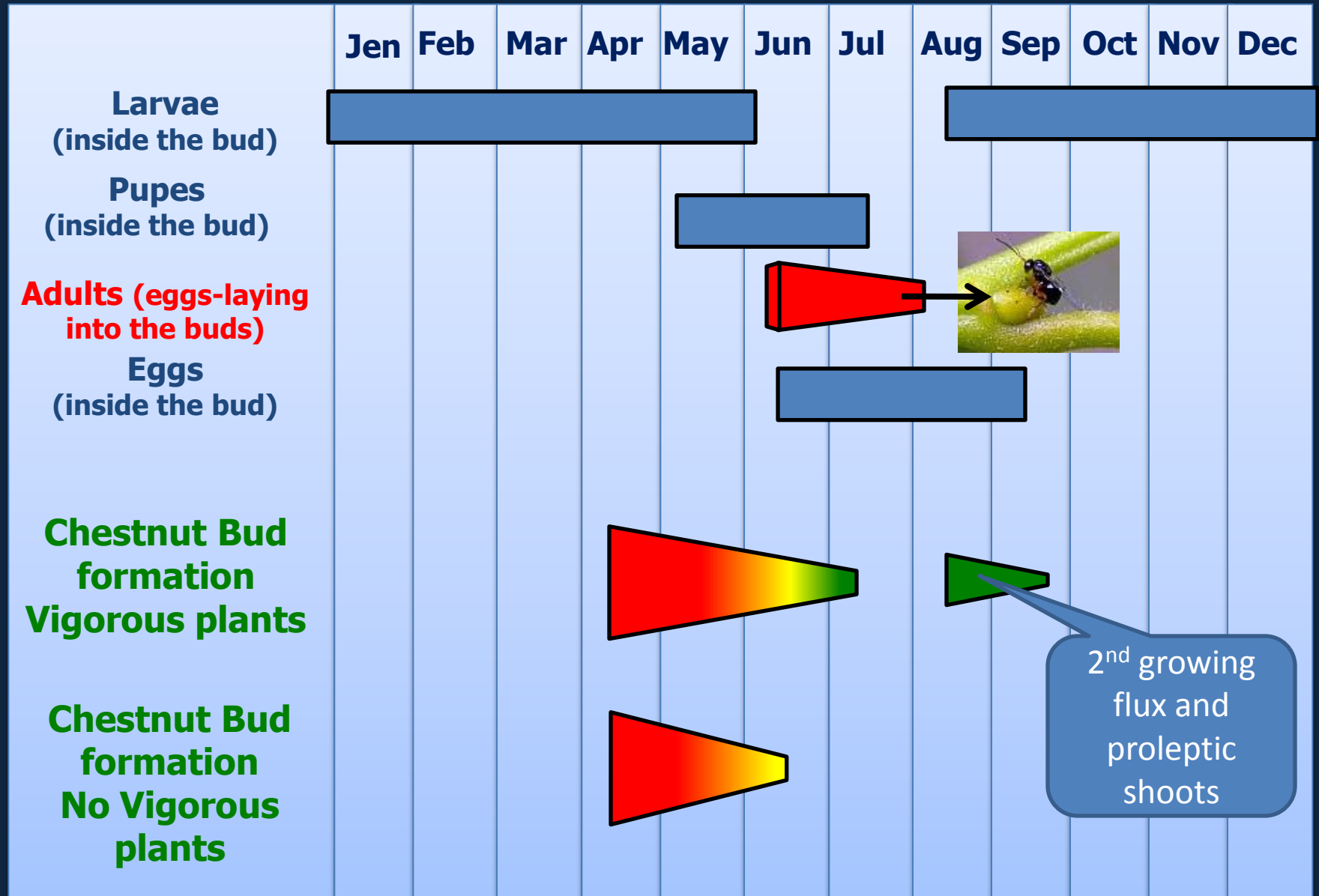
Vigorous plants tend to have  
**well developed** and **healthy**  
(or slightly damaged)  
growing **shoots** in apical  
position

Severely  
damaged shoots

(Maltoni et al., 2012)

# Results interpretation

## Time of bud formation has effects on damage susceptibility



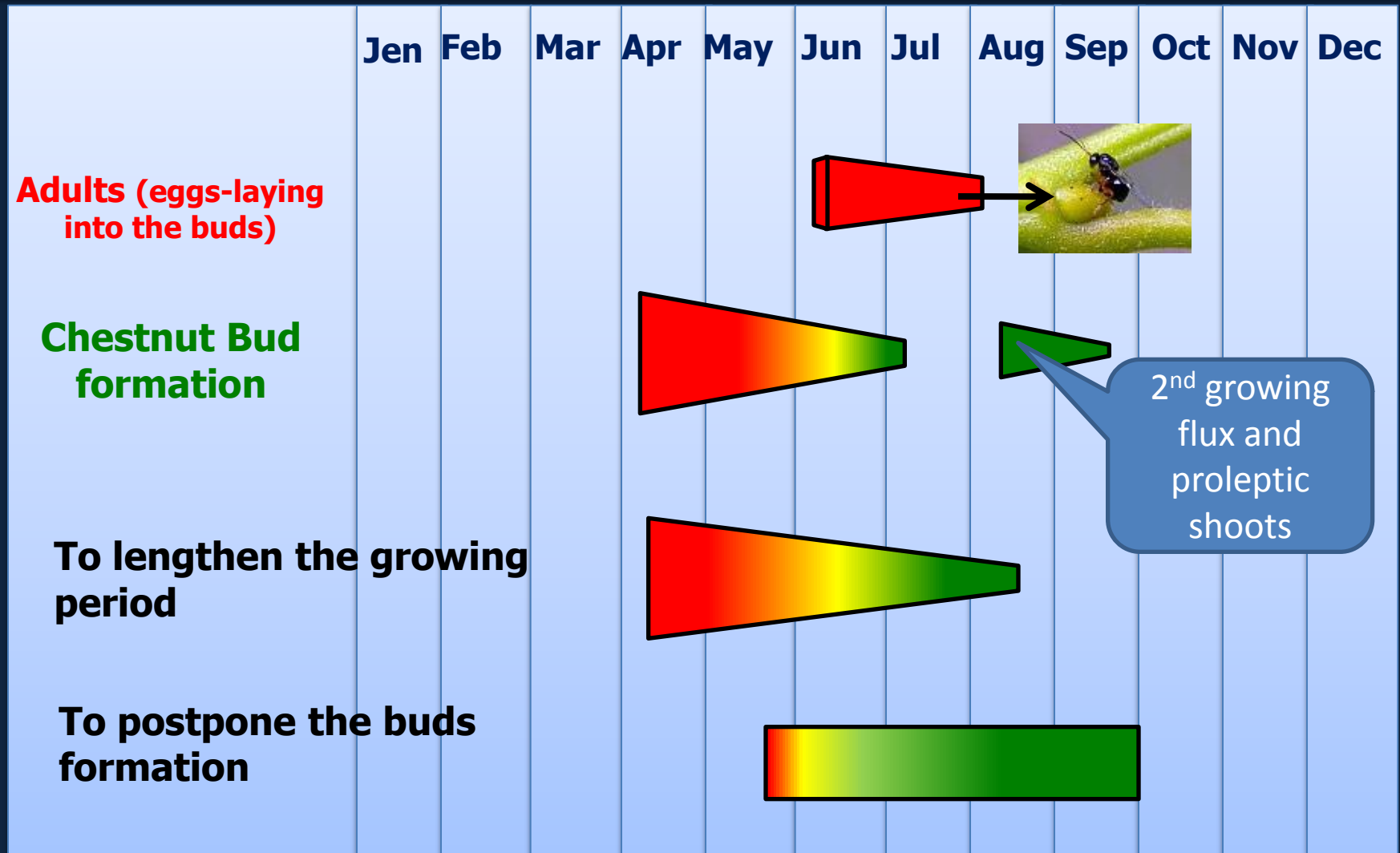


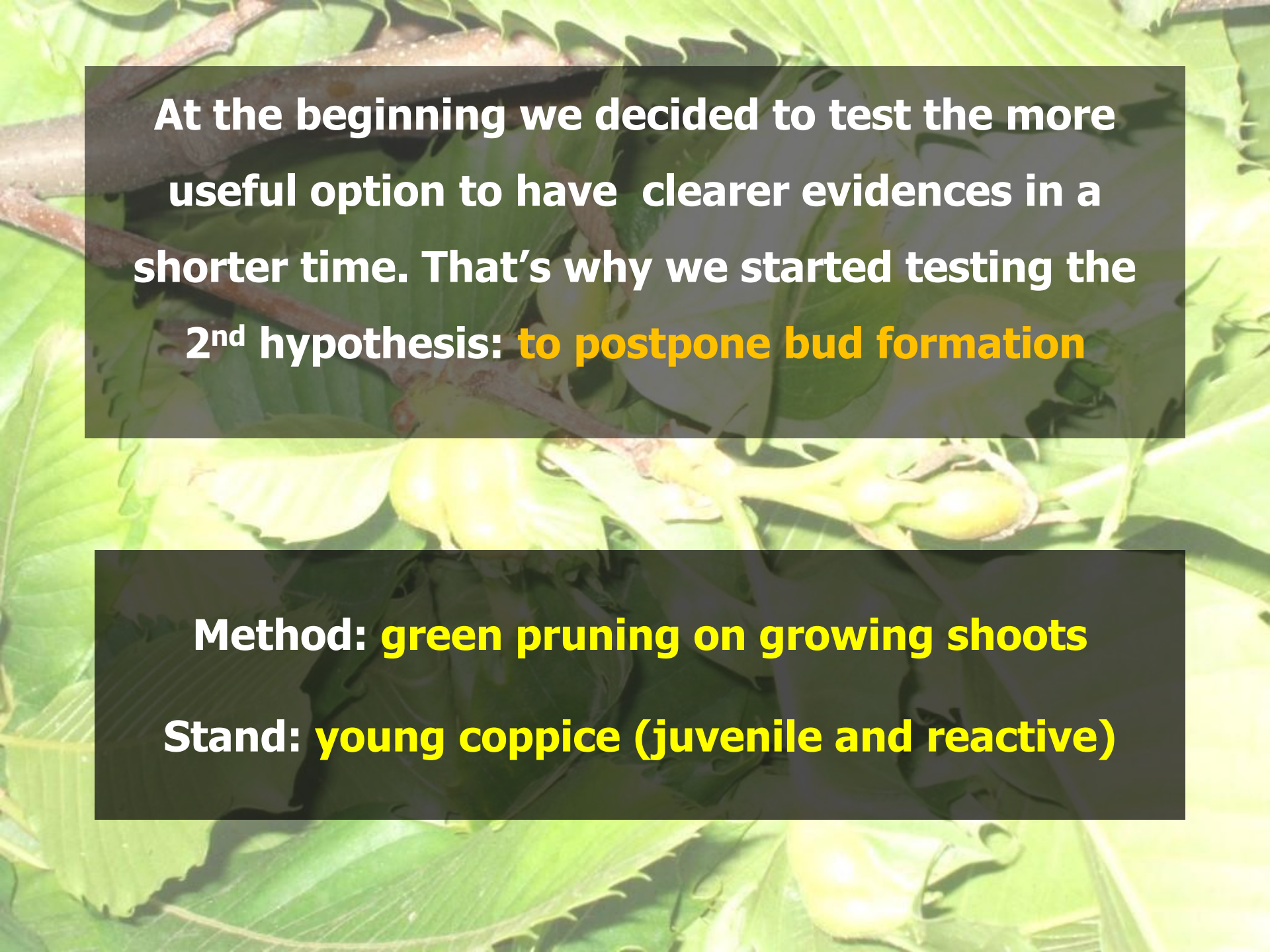
## Hypothesis:

Two possible strategies to reduce the damage postponing buds formation

1) Promote the development of vigorous spring shoots

2) Postpone shoot formation





**At the beginning we decided to test the more useful option to have clearer evidences in a shorter time. That's why we started testing the 2<sup>nd</sup> hypothesis: to postpone bud formation**

**Method: green pruning on growing shoots**  
**Stand: young coppice (juvenile and reactive)**

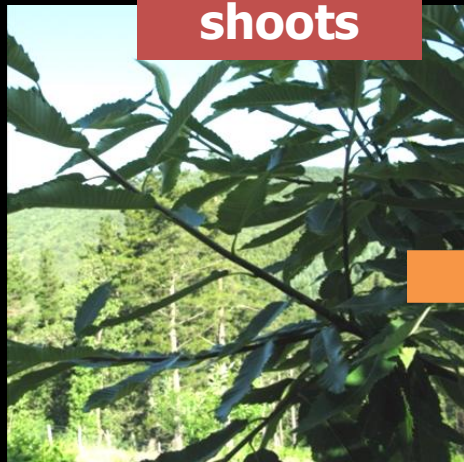


# MATERIALS and METHODS

## 2 different pruning techniques: Short-cut (SC) and Long-cut (LC)



1 year  
shoots



## 4 different pruning times (2010)

- the second half of May (M)
- the second half of June (Jn)
- the middle of July (Jl)
- the second half of August (A)





- ✚ **Central Italy (Tuscany)**
- ✚ **44°07''03''N, 10°04'58''E**
- ✚ **660 m a.s.l. , West faced**
- ✚ **Climate: sub-oceanic Mediterranean**
- ✚ **6 years pure chestnut coppice**
- ✚ **Incidence of D.k. 100%**

- ✚ **-63 sprouts (53 pruned and 10 as control)**
- ✚ **-240 shoots were pruned, 126 SC and 114 LC; 60 as control**

### **Data:**

- ✓ **Nodes number**
- ✓ **Development of shoots (presence, number, length, new nodes number, ..)**
- ✓ **Healthy nodes**
- ✓ **Heavily damaged nodes**
- ✓ **Slightly damaged nodes**

### **On**

- **The remaining shoot**
  - **Shoots grown in 2010**
  - **Shoots grown in 2011**
- Total nodes: 2794**



## ***Key points.....***



**Is chestnut able to develop new induced shoots in the pruned growing season?**

**How many shoots grew after pruning?**

**How much do the new shoots contribute to develop the crown?**



**What kind of pruning technique was the most effective?**



**Which was the most profitable pruning time?**



**Which is the best combination between pruning time and pruning technique?**

***.. Results at the end of season 2010***

# Is chestnut able to develop new induced shoots in the pruning growing season?

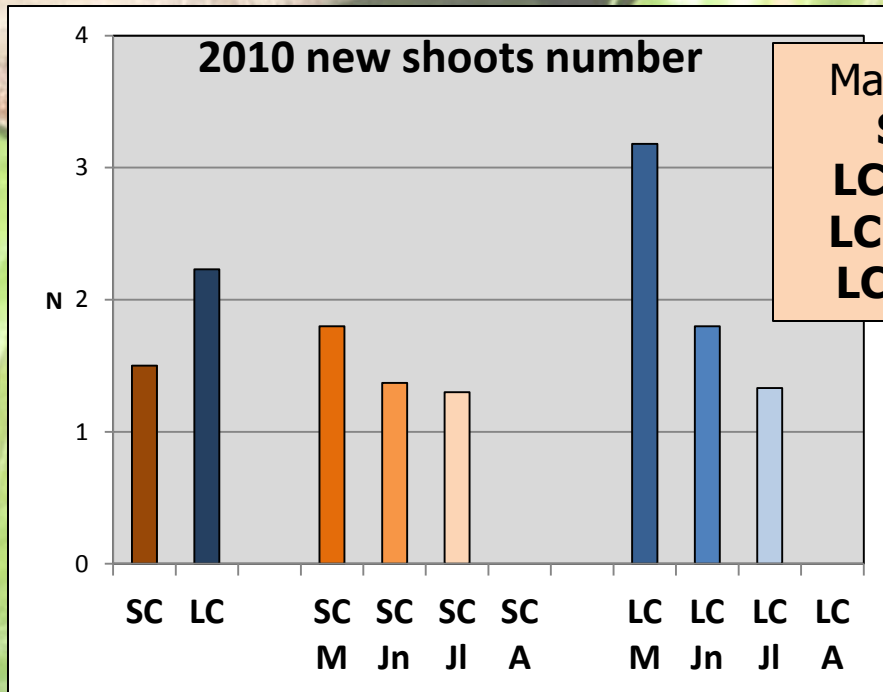
	PRUNING TECHNIQUE		
	LC	SC	
	%	%	
<b>May</b>	100	88.2	$\chi^2 = 2.6$
<b>June</b>	86.2	73.1	$\chi^2 = 1.6$
<b>July</b>	20.0	37.0	$\chi^2 = 2.1$
<b>August</b>	0	0	
	$\chi^2 = 40.5^{**}$	$\chi^2 = 13.6^{**}$	
	$M_{vs}Jn \chi^2 = 3.2$	$M_{vs}Jn \chi^2 = 1.7$	
	$M_{vs}Jl \chi^2 = 27.9^{**}$	$M_{vs}Jl \chi^2 = 11.3^{**}$	
	$Jn_{vs}Jl \chi^2 = 26.0^{**}$	$Jn_{vs}Jl \chi^2 = 7.0^{**}$	

Presence of 2010 new shoots per cut





# Is chestnut able to develop enough new induced shoots in the pruning growing season?



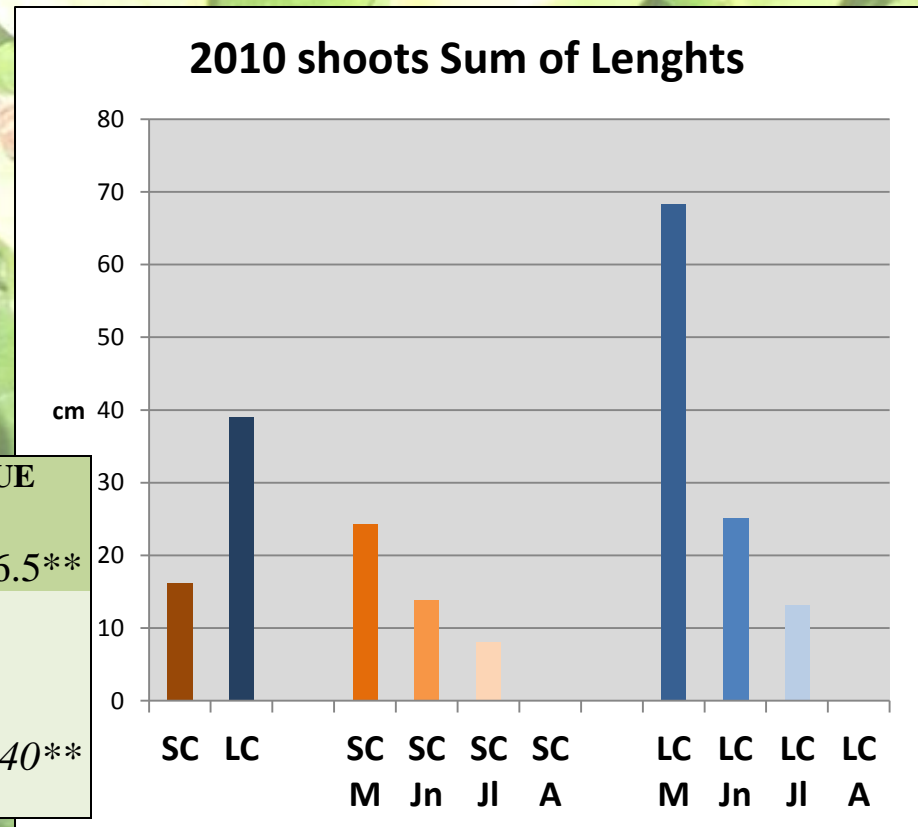
Mann-Whitney U Test:

**SC ≠ LC**  $p < 0.01$

**LC M ≠ SC M**  $p < 0.01$

**LC M ≠ LC Jn**  $p < 0.01$

**LC M ≠ LC JI**  $p < 0.01$






Multifactorial ANOVA			PRUNING TECHNIQUE		
			LC	SC	
			38.9 B	16.1 A	F=16.5**
Pruning time	M	47.7 B	68.3 b	24.3 a	F=6.40**
	Jn	20.2 A	25.1 a	13.8a	
	Jl	10.0 A	13.1 a	8.0 a	
		F=15.5**			



## *Key points.....*

 **How effective was the pruning method in limiting the attack?**

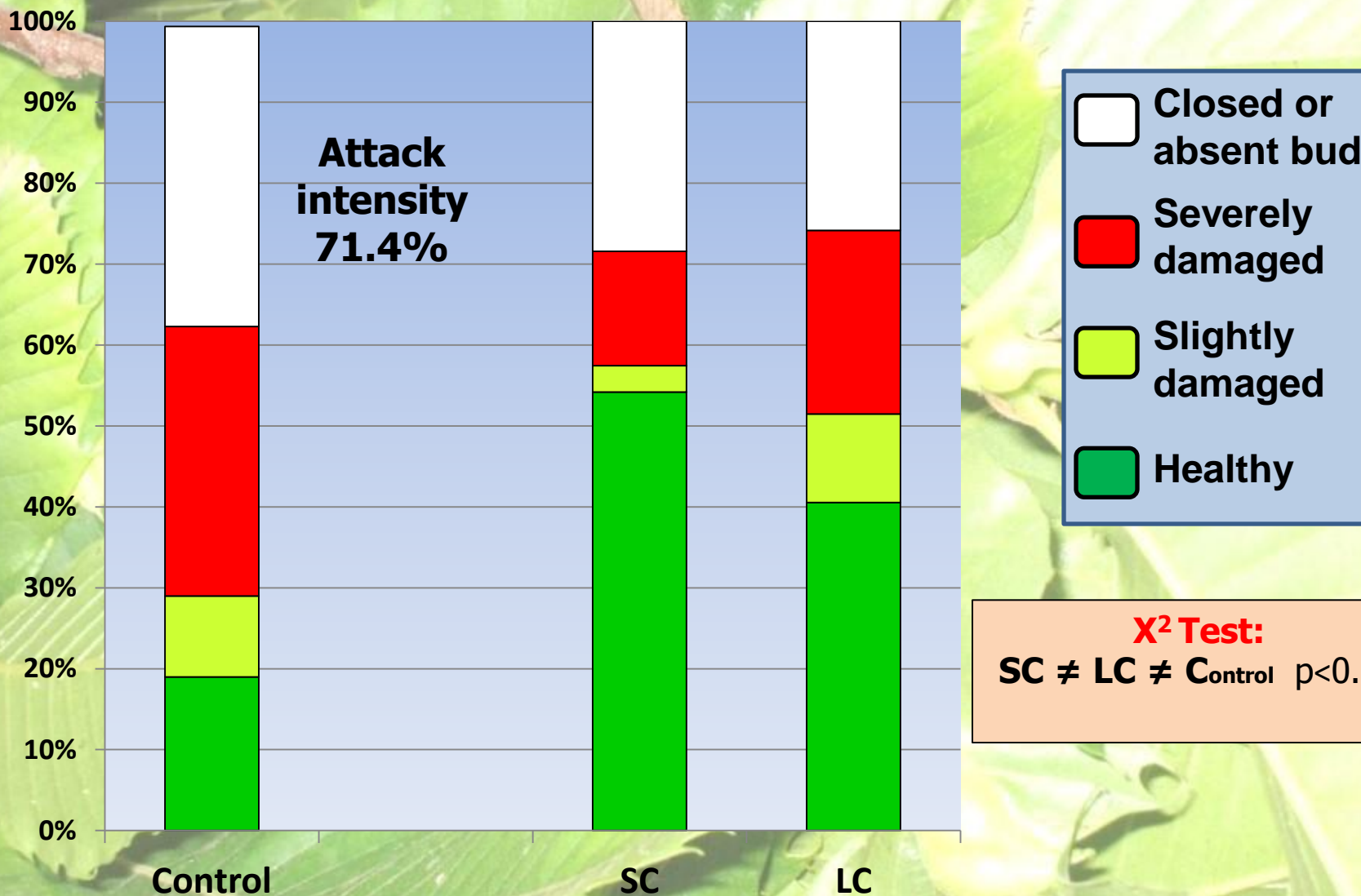
-  **Which was the most profitable pruning time to postpone bud formation?**
-  **Does the pruning technique have a significant role?**
-  **Which is the best combination between pruning time and pruning technique?**

 **What would have happened if the plants hadn't been pruned?**

***.. Results at the end of season 2011***

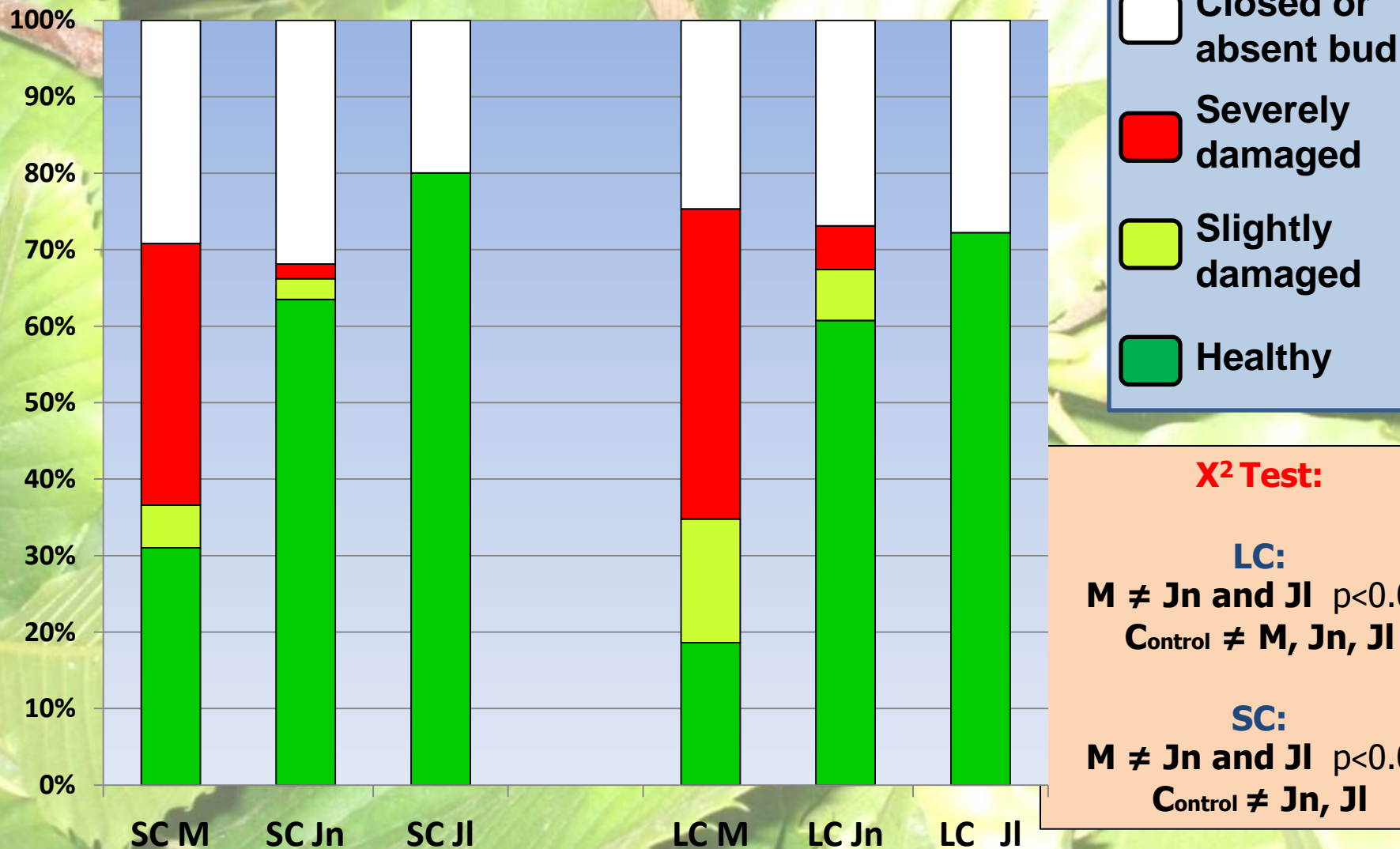
# How effective was the pruning method in limiting the attack?

## 2011 phytosanitary state of the nodes



# How effective was the pruning method in limiting the attack?

2011 phytosanitary state of the nodes



**X<sup>2</sup> Test:**

**LC:**

**M ≠ Jn and JI**  $p < 0.01$

**Control ≠ M, Jn, JI**

**SC:**

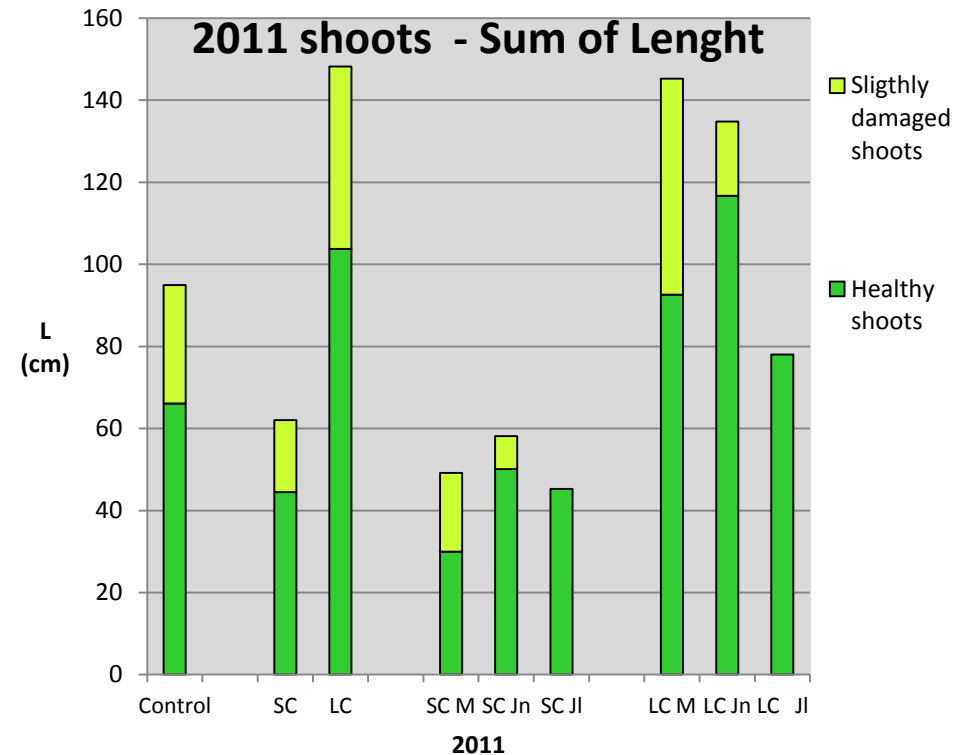
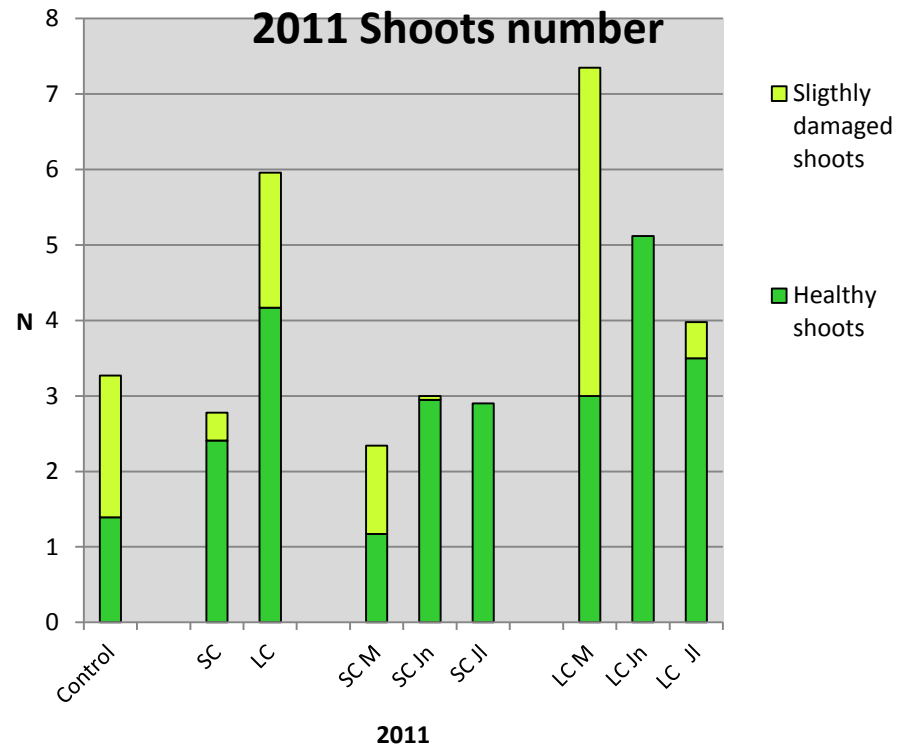
**M ≠ Jn and JI**  $p < 0.01$

**Control ≠ Jn, JI**



# Which was the most profitable pruning time to postpone bud formation?

## Does the pruning technique have a significant role?



**Mann-Whitney U Test:**  
 $p < 0.05$  in all cases except SC  
 in M and Control

**LC  $\neq$  Control (HS+SDS)**

**Pruning  
technique**

**LC**

**SC**

**Control**

**Sum of healthy shoots Length  
(cm)**

103.8 B

44.5 A

66.1 A

F= 7.30\*\*



On young grafts or in  
intensively cultivated orchards



Traditional chestnut orchards  
(most spread)

To transfer the results in **traditional chestnut orchards** is difficult because of:

- lower reactivity of mature trees
- green pruning on 1 yr growing shoots in is not practicable and unaffordable on mature trees

**The “postponing” strategy is not suitable**





**That's why we are testing the 1<sup>st</sup> hypothesis:**  
**to lengthen the growing period**  
**inducing the development of vigorous spring**  
**shoots**

**Method: pruning (winter and green) on branches**  
**Stand: old traditional chestnut orchards**



## Possible criteria

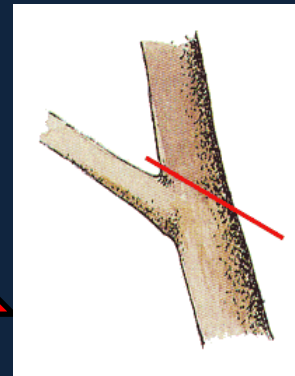
Healthy shoots from  
dormant buds



healthy but too juvenile so  
not productive (fruit)



To reinvigorate branches  
to obtain more vigorous  
spring shoots



Adopted technique

## Test on mature trees: preliminary results

### **Pruning times:**

**Winter + May, June, July** (green pruning on woody organs)

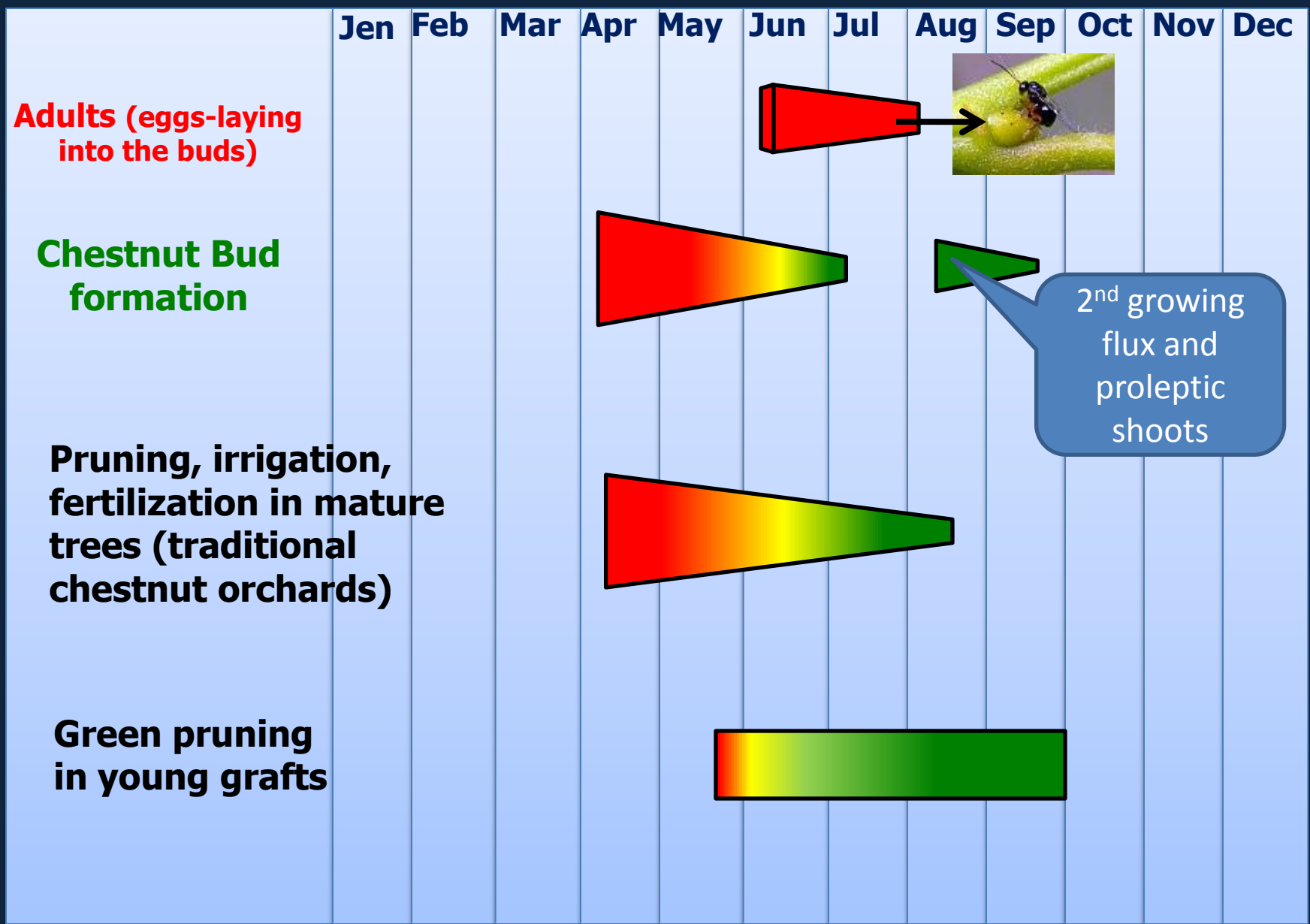
**Damage susceptibility:** not statistically significant results between pruned and non pruned plants (low presence of gall wasp?)

**Spring shoots development:** higher in pruned plants (but not statistically)

**Fruit production:** no reduction in quantity and bigger nut size in pruned plants (but not statistically different)



# Strategies and cultural practices





**It is possible to induce new healthy shoots by green pruning. Time plays an important role.**

**The best technique was the LC. The most effective time in obtaining new vigorous shoots is not the most profitable in terms of limiting damages. LC in second half of June is the best combination**

**The tested method (shortening cuts on 1 years old shoots) is suitable for young plants, as young grafts in orchards.**

**This method can interact negatively with the early stage of introduction of *Torymus sinensis*, so (in this phase) it have to be used in areas far from the introduction zones**





Test on mature trees are in progress

Preliminary results of pruning tests are positive but not significant in reducing damages

Pruning is necessary

Considering that any **environmental condition** or **cultural practice** that **lengthens** the plant growth season improves the phytosanitary state (reduce damages), it will be important to test the best effective combination of different cultural practices (pruning + fertilization and/or irrigation)

More studies are necessary

**Thank you!**

**barbara.mariotti@unifi.it**



**We want to say thank you to Italian chestnut associations and chestnut growers that helped us to start and carry on this study**



# Case study of a new method for the classification and analysis of *Dryocosmus kuriphilus* Yasumatsu damage to young chestnut sprouts

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## Pruning methods to restore *Castanea sativa* stands attacked by *Dryocosmus kuriphilus*

Alberto Maltoni • Barbara Mariotti • Douglass F. Jacobs •  
Andrea Tani